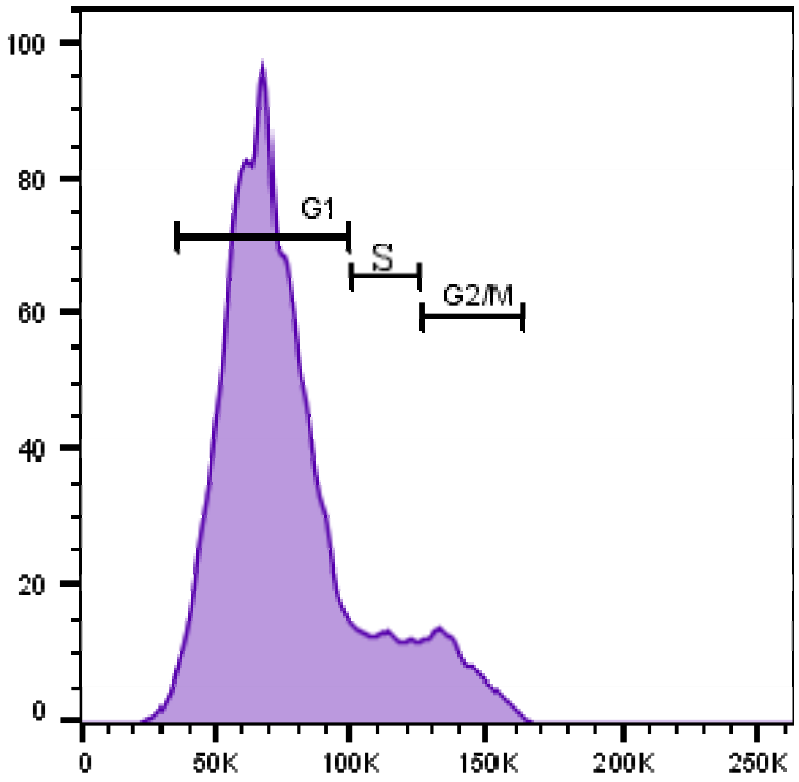
Helix NP™ Blue



C57BL/6 mouse thymocytes were fixed and permeabilized with 70% ethanol. Cells were stained with Helix NP™ Blue at 1nM.

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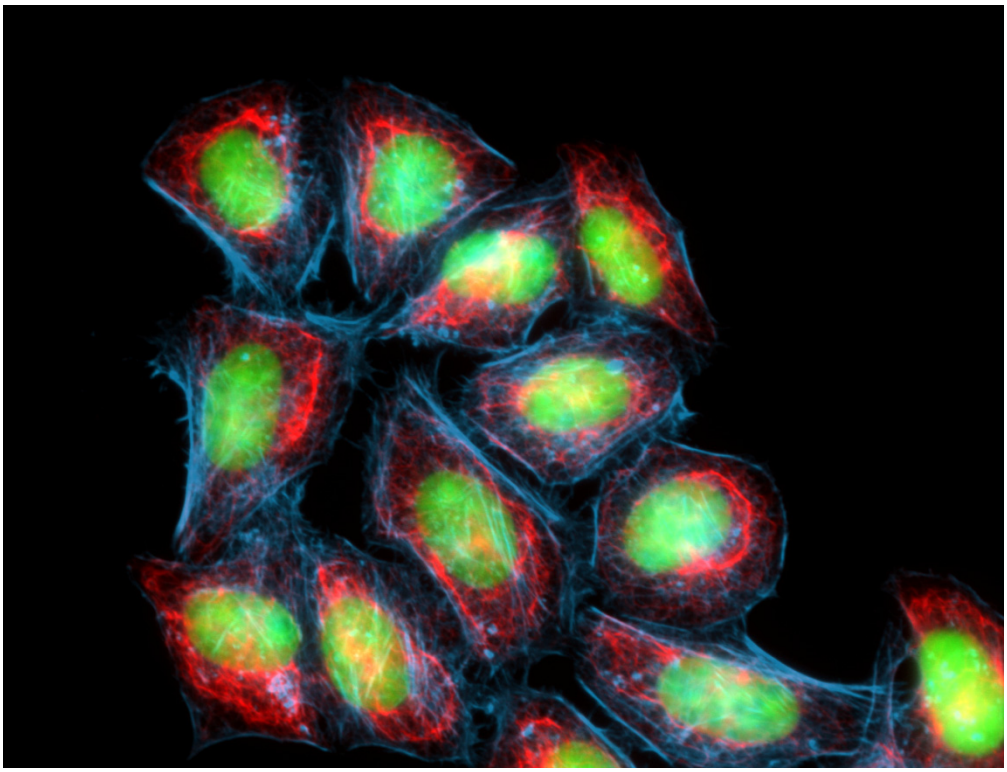
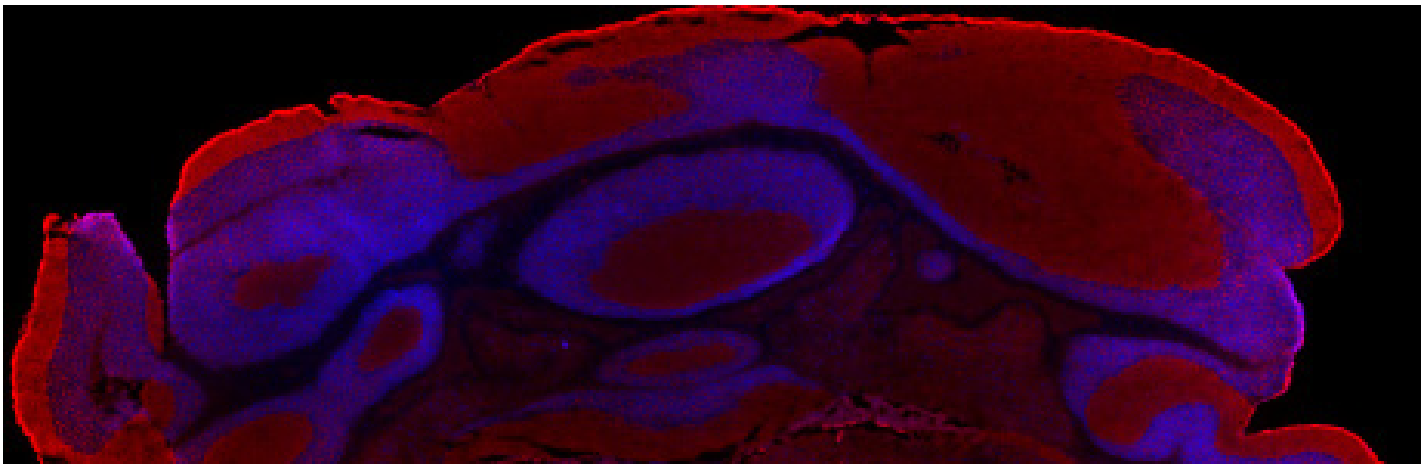


HeLa cells were fixed with 1% paraformaldehyde (PFA) for 10 minutes, permeabilized with 0.5% Triton X-100 for 10 minutes, and blocked with 5% FBS for 30 minutes. Then the cells were intracellularly stained with 5 µg/mL of Alexa Fluor® 647 anti-Cytokeratin (pan reactive) (clone C-11) antibody (red) in blocking buffer overnight followed by 25 µM of Helix NP™ Blue (green) and Flash Phalloidin™ Red 594 (blue) staining for 15 minutes at room temperature. The image was captured with a 60X objective using the Alexa Fluor® 488 filter.

Helix NP™ Blue



Mouse frozen cerebellum tissue was fixed with 4% paraformaldehyde (PFA) for 10 minutes, permeabilized with 0.5% Triton X-100 for 10 minutes, and blocked with 5% FBS for one hour. Then the tissue was intracellularly stained with 5 µM of Helix NP™ Blue (blue) 15 minutes at room temperature and co-stained with Flash Phalloidin™ Red (red). The image was captured with a 10X objective using the Alexa Fluor® 488 filter.