

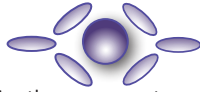


Enabling Legendary Discovery™

# Human ST2

## ELISA MAX™ Deluxe Set

Cat. No. 445004



BioLegend's ELISA MAX™ Deluxe Set contains the components necessary for the accurate quantification of natural and recombinant Human ST2. The set is designed for cost-effective and accurate quantification of Human ST2 in cell culture supernatant, serum, plasma, CSF samples or other biological fluids. BioLegend's ELISA MAX™ Deluxe Sets are sensitive, accurate, and robust.

**It is highly recommended that this instruction sheet be read in its entirety before using this product. Do not use this set beyond the expiration date.**

### Materials Provided

- Human ST2 ELISA MAX™ Capture Antibody (200X)
- Human ST2 ELISA MAX™ Detection Antibody (200X)
- Human ST2 Standard
- Avidin-HRP (1000X)
- Substrate Solution F
- Coating Buffer B (5X)
- Blocking Buffer A (2X)
- Assay Diluent B (5X)
- Assay Diluent C
- Assay Diluent E

### Introduction

IL1r1 (also known as ST2) is a member of the IL-1 superfamily, and its only known ligand is IL-33. ST2 exists in two forms as splice variants. A soluble form (sST2), which acts as a decoy receptor, sequesters free IL-33, and does not signal and a membrane-bound form (ST2), which activates the MyD88/NF-κB signaling pathway to enhance mast cell, Th2, regulatory T cell (Treg), and innate lymphoid cell type 2 functions. sST2 levels are increased in patients with intestinal disease, pulmonary disease, heart disease and various other inflammatory diseases.

### Principle of the Test

BioLegend's ELISA MAX™ Deluxe Set is a sandwich Enzyme-Linked Immunosorbent Assay (ELISA). A Human ST2 specific monoclonal antibody is first coated on a 96-well plate. Standards and samples are added to the wells, and ST2 binds to the immobilized capture antibody. Next, a biotinylated anti-human ST2 detection antibody is added, producing an antibody-antigen-antibody "sandwich". Avidin-horseradish peroxidase is subsequently added, followed by TMB Substrate Solution, producing a blue color in proportion to the concentration of ST2 present in the sample. Finally, the Stop Solution changes the reaction color from blue to yellow, and the microwell absorbance is read at 450 nm with a microplate reader.

For research purposes only. Not for use in diagnostic or therapeutic procedures.

### Materials to be Provided by the End-User

- Microwell plates: BioLegend Cat. No. 423501 is recommended
- Wash Buffer: BioLegend Cat. No. 421601 is recommended, or PBS + 0.05% Tween-20
- Stop Solution: BioLegend Cat. No. 423001 is recommended, or acid solution, e.g. 2N H<sub>2</sub>SO<sub>4</sub>
- Plate Sealers: BioLegend Cat. No. 423601 is recommended
- PBS (Phosphate-Buffered Saline): 8.0 g NaCl, 1.16 g Na<sub>2</sub>HPO<sub>4</sub>, 0.2 g KH<sub>2</sub>PO<sub>4</sub>, 0.2 g KCl, add deionized water to 1 L; pH to 7.4, 0.2 μm filtered
- Deionized (DI) water
- A microplate reader capable of measuring absorbance at 450 nm
- Adjustable pipettes to measure volumes ranging from 2 μL to 1 mL
- Wash bottle or automated microplate washer
- Log-Log graph paper or software for data analysis
- Tubes to prepare standard dilutions
- Timer

### Storage Information

- Store kit components between 2°C and 8°C.
- After reconstitution of the lyophilized standard with 1X Assay Diluent B, aliquot into polypropylene vials and store at -70°C for up to one month. Avoid repeated freeze/thaw cycles.
- Prior to use, bring all components to room temperature (18°C-25°C). Upon assay completion return all components to appropriate storage conditions.

### Health Hazard Warnings

- Reagents that contain preservatives may be harmful if ingested, inhaled or absorbed through the skin. Refer to the MSDS online for details ([www.biolegend.com/support/#msds](http://www.biolegend.com/support/#msds)).
- Substrate Solution is harmful if ingested. Additionally, avoid skin, eye or clothing contact.
- To reduce the likelihood of blood-borne transmission of infectious agents, handle all serum and/or plasma in accordance with NCCLS regulations.

### Specimen Collection and Handling

**Cell Culture Supernatant:** If necessary, centrifuge to remove debris prior to analysis. Samples can be stored at < -20°C. Avoid repeated freeze/thaw cycles.

**Serum:** Use a serum separator tube and allow clotting for at least 30 minutes, then centrifuge for 10 minutes at 1,000 X g. Remove serum layer and assay immediately or store serum samples at < -20°C. Avoid repeated freeze/thaw cycles. Serum specimens should be clear and non-hemolyzed.

**Plasma:** Collect blood sample in a citrate, heparin or EDTA containing tube. Centrifuge for 10 minutes at 1,000 X g within 30 minutes of collection. Assay immediately or store plasma samples at < -20°C. Avoid repeated freeze/thaw cycles. Plasma specimens should be clear and non-hemolyzed.

### Reagent and Sample Preparation

**Do not mix reagents from different sets or lots. Reagents and/or antibodies from different manufacturers should not be used with this set. All reagents should be diluted immediately prior to use.**

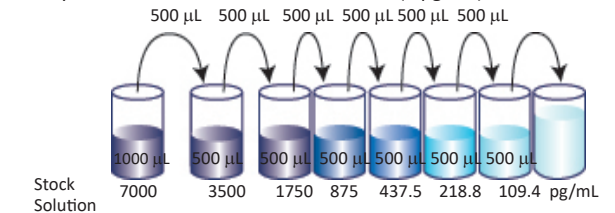
**NOTE: Precipitation of 5X Assay Diluent B may be observed when stored long term between 2°C and 8°C. The precipitation does not alter the performance of the buffer. If heavy precipitation is observed after the dilution to 1X Assay Diluent B, it can be filtered to clarify the solution.**

### Preparation of 1X Reagent for 1 Plate

Material	Dilute with
2.4 mL of Coating Buffer B (5X)	9.6 mL of Deionized Water
60 μL of Capture Antibody (200X)	12 mL of 1X Coating Buffer B
12 mL of Assay Diluent B (5X)	48 mL of PBS
12 mL of Blocking Buffer A (2X)	12 mL of Deionized Water
60 μL of Detection Antibody (200X)	12 mL of 1X Assay Diluent B
12 μL of Avidin-HRP (1,000X)	12 mL of 1X Assay Diluent B

Lyophilized vials are under vacuum pressure. Reconstitute lyophilized standard with 1 mL of Assay Diluent B. Allow the reconstituted standard to sit for 15 minutes at room temperature, then mix gently prior to making dilutions.

Prior to use, prepare 1,000 μL of the top standard at a concentration of 7000 pg/mL from the stock solution in Assay Diluent B (refer to **Lot-Specific Certificate of Analysis/ELISA MAX™ Deluxe Set Protocol**). Perform six two-fold serial dilutions of the 7000 pg/mL top standard with Assay Diluent B in separate tubes. After diluting, the human ST2 standard concentrations are 7000 pg/mL, 3500 pg/mL, 1750 pg/mL, 875 pg/mL, 437.5 pg/mL, 218.8 pg/mL, and 109.4 pg/mL, respectively. Assay Diluent B serves as the zero standard (0 pg/mL).



Samples: For CSF samples, the end user may need to determine the dilution factors in a preliminary experiment. In general, a 2-fold dilution in Assay Diluent B is recommended for serum, plasma and cell culture supernatant samples. All samples can be diluted further in Assay Diluent B to fit within the range of the assay as determined by the end user. Either Assay Diluent C will be added to the wells first to measure plasma, cell culture supernatant and CSF samples or Assay Diluent E will be added first to measure serum samples.

### Assay Procedure

**Do not use sodium azide in any solutions as it inhibits the activity of the horseradish-peroxidase enzyme.**

- One day prior to running the ELISA, dilute Capture Antibody in 1X Coating Buffer as described in Reagent Preparation. Add 100 μL of this Capture Antibody solution to all wells of a 96-well plate. Seal plate and incubate overnight (16-18 hrs) between 2°C and 8°C.
- Bring all reagents to room temperature (RT) prior to use. It is strongly recommended that all standards and samples be run in duplicate or triplicate. A standard curve is required for each assay.
- Wash plate 4 times with at least 300 μL Wash Buffer per well and blot residual buffer by firmly tapping plate upside down on absorbent paper. **All subsequent washes should be performed similarly.**
- To block non-specific binding and reduce background, add 200 μL 1X Blocking Buffer A per well.
- Seal plate and incubate at RT for 1 hour with shaking on a plate shaker (e.g. 200 rpm with a 0.3 cm circular orbit). All subsequent incubation with shaking should be performed similarly.

6. While plate is being blocked, prepare the appropriate sample dilutions (if necessary) and standards.

**7. For measuring cell culture medium, plasma and CSF samples:**

- Wash plate 4 times with Wash Buffer.
- Add 50 µL/well of Assay Diluent E to the standard and sample wells.
- Add 50 µL/well of standards prepared above to the standard wells. Add 50 µL/well of cell culture medium, plasma or CSF samples to the sample wells. If dilution is required, samples should be diluted in Assay Diluent B before adding to the wells.

**For measuring serum samples:**

- Wash plate 4 times with Wash Buffer.
- Add 50 µL/well of Assay Diluent C to the standard and sample wells.
- Add 50 µL/well of standards prepared above to the standard wells. Add 50 µL/well of serum samples to the sample wells. If dilution is required, samples should be diluted in Assay Diluent B before adding to the wells.

8. Seal plate and incubate at RT for 2 hours with shaking.

9. Wash plate 4 times with Wash Buffer.

10. Add 100 µL of diluted Detection Antibody solution to each well, seal plate and incubate at RT for 1 hour with shaking.

11. Wash plate 4 times with Wash Buffer.

12. Add 100 µL of diluted Avidin-HRP solution to each well.

13. Seal plate and incubate at RT for 30 minutes with shaking.

14. Wash plate 5 times with Wash Buffer. For this final wash, soak wells in Wash Buffer for 30 seconds to 1 minute for each wash. This will help minimize background.

15. Add 100 µL of TMB Substrate Solution F and incubate **in the dark** for 10 minutes\*. Positive wells should turn blue in color. It is not necessary to seal the plate during this step.

16. Stop reaction by adding 100 µL of Stop Solution to each well. Positive wells should turn from blue to yellow.

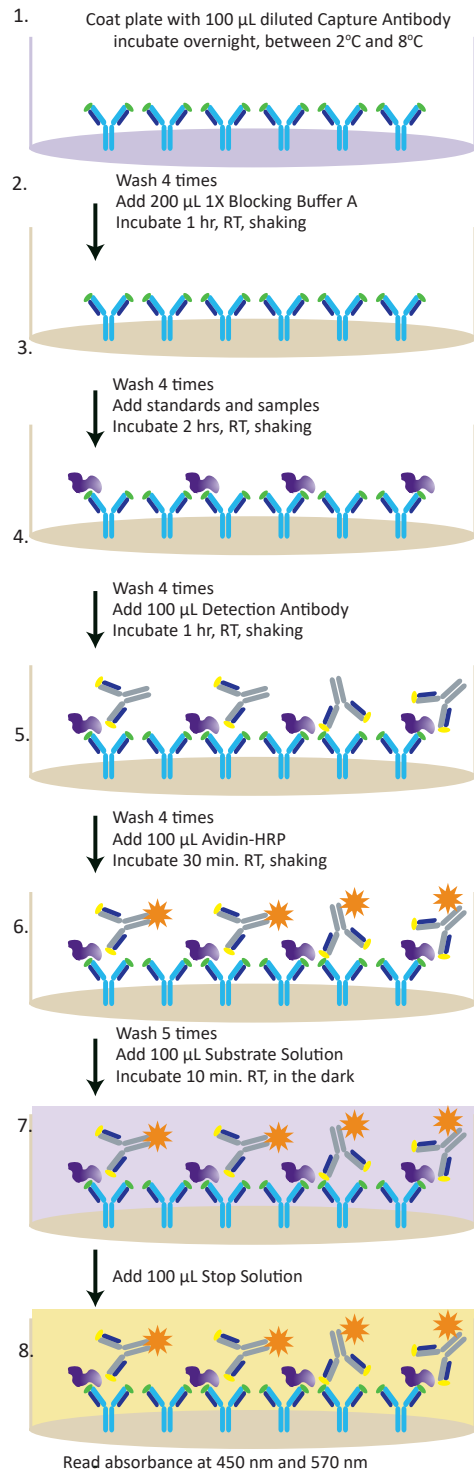
17. Read absorbance at 450 nm within 15 minutes. If the reader can read at 570 nm, the absorbance at 570 nm can be subtracted from the absorbance at 450 nm.

**\*Optimal substrate incubation time depends on laboratory conditions and the optical linear ranges of ELISA plate readers**

### Calculation of Results

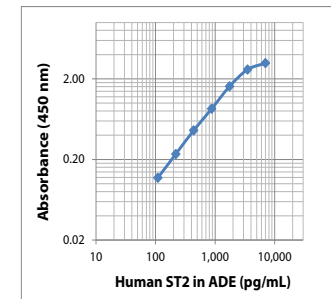
Plot the standard curve on log-log axis graph paper with analyte concentration on the x-axis and absorbance on the y-axis. Draw a best fit line through the standard points. To determine the unknown analyte concentrations in the samples, find the absorbance value of the unknown on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the corresponding analyte concentration. If the samples were diluted, multiply by the appropriate dilution factor. The data is best calculated with computer-based curve-fitting software using a 5- or 4-parameter logistics curve-fitting algorithm. If a test sample's absorbance value falls outside the standard curve ranges, that test sample needs to be reanalyzed at a higher or lower dilution as appropriate.

### Assay Procedure Summary



### Typical Data

**Standard Curve:** This standard curve is generated at BioLegend for demonstration purposes only. A standard curve must be run with each assay.



### Performance Characteristics

**Sensitivity:** The minimum detectable concentration of Human ST2 for this set in ADC is 45.14 pg/mL and the minimum detectable concentration of Human ST2 for this set in ADE is 48.75 pg/mL

**Specificity:** No significant cross reactivity was observed when this kit was tested on multiple human and mouse recombinant proteins. Interference was observed at 780 pg/mL of human IL-33, suggesting the assay recognizes free ST2.

### Troubleshooting

Refer to the troubleshooting guidelines at BioLegend's website for details ([http://www.biolegend.com/elisa\\_troubleshooting](http://www.biolegend.com/elisa_troubleshooting)).

### BioLegend, Inc.

BioLegend is ISO 13485 certified.  
8999 BioLegend Way, San Diego, CA 92121  
Tel: 1-858-768-5800, Fax: 1-877-455-9587  
[www.biolegend.com](http://www.biolegend.com)



For other technical resources, please visit:  
[www.biolegend.com/support](http://www.biolegend.com/support) or  
email: [techserv@biolegend.com](mailto:techserv@biolegend.com)